Urinary levels of intact, free beta and beta core fragment of human chorionic gonadotrophin (hCG) in early pregnancy John Walter Larsen¹, Saji Eapen², Sarah Johnson², Lorrae Marriott^{2,} Michael J. Zinaman³

Introduction

- Home and laboratory pregnancy testing relies on the detection of human chorionic gonadotrophin (hCG),¹ a glycoprotein with two non-covalently linked subunits: alpha (hCG- α) and beta (hCG-B)
- Multiple forms of hCG are present in the serum and urine of pregnant women: as shown in Figure 1
- The degradation product, B-core-hCG is only present in urine and becomes the predominate form detectable in later pregnancy.



- Reference ranges have been published for intact urinary hCG, but not for other forms present in the urine of pregnant women^{1,3}
- Free-B-hCG and B-core-hCG have a different profile of daily rise compared with intact hCG
- Very high levels of B-core-hCG, that can occur in later pregnancy, have caused false negative point-of-care pregnancy test results which can have serious clinical consequences
- Although laboratory testing for B-core-hCG interference has been conducted on both home and point-of-care pregnancy tests, there is some debate as to what the most appropriate testing methodology should be^{4,5}
- The ratio of intact: free-B-hCG has been related to pregnancy viability⁶
- Therefore, it is important to obtain further understanding of the ranges of these forms of hCG in pregnancy and their relationship with the total level of hCG.

Purpose of the study

• This study sought to improve the understanding of the levels of free-B and B-core-hCG in viable pregnancies.



Methods

- pregnancies throughout early pregnancy
- enabled accurate assignment of pregnancy duration for each volunteer
- Perkin Elmer assay for intact hCG and free-B assays.
- Mean and standard deviations (SD) by the day of pregnancy were derived.

Results

first trimester of pregnancy



Figure 2: Intact hCG, free B-hCG (A) and B-core-hCG (B) concentrations during the first trimester of pregnancy

- However, free B-hCG was not consistently detectable in urine until day 21
- intact hCG
- B-core-hCG had a different profile, appearing in urine later than intact hCG (day 19), yet becoming the predominant form by day 35
- High levels of B-core-hCG were only present when there was also intact hCG in the sample
- was 10,003 pmol/l (4017 mlU/ml), as shown in Figure 3.

1 The George Washington University, Washington, District of Columbia, USA. 2 SPD Development Company Ltd., Bedford, UK. 3 Albert Einstein College of Medicine, Jacobi Medical Center, Bronx, New York, USA.

Daily early morning urine samples were collected from women with viable

• The samples were collected pre-conception to enable the day of ovulation to be determined for each woman by the detection of the LH surge (AutoDELFIA quantitative LH assay, with ovulation presumed as LH surge + 1 day). This Intact, free-B and B-core-hCG were measured using AutoDELFIA immunoassays, using in-house reagents for B-core-hCG assays, and the



• As expected, intact hCG was present in the urine of pregnant women 8 days following ovulation, and showed a consistent rise throughout early pregnancy • Free B-hCG appeared in urine at a constant ratio of approximately 1:100 of

• The minimum level of intact hCG in samples with B-core-hCG >500,000 pmol/l



References

- 5. Gronowski AM, et al. Clin Chem. (2009) 55: 1885–1886.

Declaration of Interest

This study was funded by SPD Development Company Ltd., a wholly owned subsidiary of SPD Swiss Precision Diagnostics GmbH. Sarah Johnson, Saji Eapen and Lorrae Marriot are employees of SPD Development Company Ltd.

cause false negatives in some pregnancy tests

• As these high levels of *B*-core-hCG always occur in the presence of total hCG, testing for interference of assays by B-core-hCG should be conducted using samples that also contain intact hCG • Only assays that demonstrate they are unaffected by B-core-hCG interference should be used in later pregnancy.

Gnoth C and Johnson J. Geburtsh Frauenheilk. (2014) 74: 661–669. 2. Birstow A, et al. Clin Chem. (2005) 51: 177–182.

3. Larsen J, et al. Gynecol Obstet. (2013) 123: 189–195.

. Gronowski AM. Clin Chem. (2009) 55: 1900–1904.

6. Nerenz RD and Gronowski AM. Clin Chem. (2015) 61: 483-486.

